In-vivo Animal Core (IVAC-ADL) Clinical Pathology Sample Collection Guidelines

Sample Drop-off Locations, 9:00 a.m. – 4:00 p.m.
3527 Animal Research Facility North Campus Research Complex
1150 W. Medical Center Drive 2800 Plymouth Road B36/G157
Ann Arbor, MI 48109-0614 Ann Arbor, MI 48109-2800
ARF Phone: (734) 936-3803 NCRC Phone: (734) 936-3395
IVAC Manager: (734) 647-0731 Email: ULAM-IVA.CLAB@umich.edu

• For protocol approval information, please contact the Animal Care and Use Office at ACUOffice@umich.edu.
• For technique training, please contact the ULAM Training Core at ULAM-TRAININGCORE@umich.edu or 734-763-8039.
• For information about assistance with sample collection, please contact ULAM Technical Services at ULAM-TECHSERVICES@umich.edu.
• For questions about specific volumes, please contact us at 734-936-3395 or 734-936-3803.

Minimum Volumes:
• CBC (Complete Blood Count):
  o Draw 50µl whole blood in lavender (EDTA) Microvette 100 tube
  o Other anticoagulants may interfere with results
  o Be sure to immediately and thoroughly mix sample after drawing
  o CBC samples must never be stored on ice
  o CBC samples cannot be analyzed after 24 hours

• Chemistries:
  o Chemistries are performed on serum or heparinized plasma
  o Blood may be collected in the yellow top serum separator microtainer, heprinized tubes, or any tube that does not contain additives
  o Normally, half the volume of whole blood is serum (i.e. 100 µl whole blood gives 50 µl serum
  o Allow blood to clot for at least 10 minutes, then centrifuge sample for 8-10 minutes at 7,000 rpm
  o If samples will not be run in 24 hours or more, spin and separate serum, then freeze serum at -20ºC.
  o The volume of serum required depends on the number of tests ordered. Consult the laboratory as needed

Submission Notes:
• A current account number (short code) must always be given
• Syringes with needles will NOT be accepted
• If running chemistries the only anticoagulant that is acceptable for plasma is lithium heparin
• Fecal specimens must be in proper containers (no gloves)
• Urine samples should be a clean catch collected in a sterile container (leukocyte values may be inaccurate on random catch samples)
• Microbiology samples should be submitted promptly after collection

Supplies:
***Though the ADL does not provide supplies, we do have a list of sources***

<table>
<thead>
<tr>
<th>Item</th>
<th>Usage</th>
<th>Vendor</th>
<th>Stock #</th>
</tr>
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<tbody>
<tr>
<td>Serum separator microtainer/yellow</td>
<td>Chemistries, Serology</td>
<td>Fisher</td>
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<td>Culturette collection system</td>
<td>Microbiology swabs</td>
<td>Fisher</td>
<td>B432009</td>
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<tr>
<td>EDTA Microvette 100</td>
<td>CBC</td>
<td>Fisher/Sarsdet</td>
<td>NC0416471</td>
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