

In-vivo Animal Core (IVAC-ADL) Clinical Pathology Sample Collection Guidelines

Sample Drop-off Locations, 9:00 a.m. – 4:00 p.m.

3527 Animal Research Facility
 1150 W. Medical Center Drive
 Ann Arbor, MI 48109-0614
 ARF Phone: (734) 936-3803
 IVAC Manager: (734) 647-0731

North Campus Research Complex
 2800 Plymouth Road B36/G157
 Ann Arbor, MI 48109-2800
 NCRC Phone: (734) 936-3395
 Email: ULAM-IVACLAB@umich.edu

- For protocol approval information, please contact the Animal Care and Use Office at ACUOffice@umich.edu.
- For technique training, please contact the ULAM Training Core at ULAM-TRAININGCORE@umich.edu or 734-763-8039.
- For information about assistance with sample collection, please contact ULAM Technical Services at ULAM-TECHSERVICES@umich.edu.
- For questions about specific volumes, please contact us at 734-936-3395 or 734-936-3803.

Minimum Volumes:

- **CBC (Complete Blood Count):**
 - o Draw 50µl whole blood in lavender (EDTA) Microvette 100 tube
 - o Other anticoagulants may interfere with results
 - o Be sure to immediately and thoroughly mix sample after drawing
 - o CBC samples must *never* be stored on ice
 - o CBC samples cannot be analyzed after 24 hours
- **Chemistries:**
 - o Chemistries are performed on serum or heparinized plasma
 - o Blood may be collected in the yellow top serum separator microtainer, heparinized tubes, or any tube that does not contain additives
 - o Normally, half the volume of whole blood is serum (i.e. 100 µl whole blood gives 50 µl serum)
 - o Allow blood to clot for at least 10 minutes, then centrifuge sample for 8-10 minutes at 7,000 rpm
 - o If samples will not be run in 24 hours or more, spin and separate serum, then freeze serum at -20°C.
 - o The volume of serum required depends on the number of tests ordered. Consult the laboratory as needed

Submission Notes:

- A current account number (short code) must always be given
- Syringes with needles will NOT be accepted
- If running chemistries the only anticoagulant that is acceptable for plasma is lithium heparin
- Fecal specimens must be in proper containers (no gloves)
- Urine samples should be a clean catch collected in a sterile container (leukocyte values may be inaccurate on random catch samples)
- Microbiology samples should be submitted promptly after collection

Supplies:

Though the ADL does not provide supplies, we do have a list of sources

Item	Usage	Vendor	Stock #
Serum separator microtainer/yellow	Chemistries, Serology	Fisher	02-675-184
Culturette collection system	Microbiology swabs	Fisher	B432009
EDTA Microvette 100	CBC	Fisher/ Sarsdet	NC0416471